

# Japan's comments on the proposed amendments of the Manual of Diagnostic Tests for Aquatic Animals for "Infection with Ranavirus" in the Aquatic Animal Health Standards Commission Report of the October 2010 meeting

## General Comments

In Article 4.3.1.2.2. (*Antibody-based antigen detection methods*) and Article 4.3.1.2.3. (*Molecular techniques*), the use of anti-EHNV antibodies and primers designed for EHNV is recommended for detection of amphibian ranavirus. As EHNV is distinct from amphibian ranavirus, it is an initial premise that above-mentioned antibodies and primers can be used for detection of all currently known amphibian ranaviruses.

Assuming that the initial premise is correct, in order to achieve more accurate positive results for amphibian ranavirus in tests using the above-mentioned antibodies and/or primers, Japan believes it essential to add one more step to the proposed detection process for further examining virus character, especially for confirming that the sample in question is negative for EHNV.

In addition, given that the main objective of the proposed methods for antibody-based antigen detection is to detect the viruses that are pathogenic to amphibians, Japan suggests the use of antibodies specifically targeting such amphibian ranavirus as FV3 and BIV in the future, bearing in mind that the development and the practical use of such antibodies require further time at this time.

## Specific Comments

NOTE: Please find the following specific comments in which proposed insertion is underlined and proposed deletion is ~~struck out~~. Any deletion or insertion by Japan shall be written in red on this paper.

### (Proposed draft)

#### 1. Scope

For the purpose of this chapter, ranavirus disease is considered to be systemic clinical or subclinical infection with a member of the genus *Ranavirus* with the exception of epizootic haematopoietic necrosis virus and European catfish virus.

### (Rationale)

For its consistency with Article 8.2.1. of the Aquatic Animal Health Codes.

(Proposed draft)

Table 5.1 cont. Methods for targeted surveillance and diagnosis

Method	Targeted surveillance				Presumptive diagnosis	Confirmatory diagnosis
	Ova/mil t	Tadpoles	Metamorphs	Adults		
Cell culture	na	a	a	a	a	a
Antigen-capture ELISA	na	a	a	a	b	b
Antibody-capture ELISA	na	d	d	c	c	d
PCR-REA	na	d	a	d	c	<u>a</u> <u>c</u>
PCR sequence analysis	na	d	d	d	c	<u>a</u> <u>c</u>

**(Rationale)**

In Article 4.3.1.2.3, it is recommended to use MCP-1 or MCP-2 as a primer in PCR methods. However, MCP-1 and MCP-2 are primarily designed to target EHNV. Japan concerns that the amphibian ranavirus infection may be overlooked in the proposed methods. Therefore, Japan requests OIE to replace the MCP-1 and MCP-2 with primers specifically targeting amphibian ranavirus. Otherwise, Japan proposes the change of the designation for “confirmatory diagnosis” of PCR-REA in Table 5.1 from “a” to “c”.